

Soluble Starch Synthase(SSS) Activity Assay Kit

Note: Take two or three different samples for prediction before test.

Operation Equipment: Spectrophotometer

Cat No: AK0413

Size: 50T/48S

Components:

Extract solution: Liquid 50 mL×1, store at 4°C .

Reagent I: Liquid 40 mL×1, store at 4°C .

Reagent II: Powder×2, store at 4°C .

Reagent III: Powder×2, store at -20°C .

Reagent IV: Powder×2, store at -20°C . Add 5 mL Reagent I before use.

Reagent V: Powder×2, store at -20°C . Add 8 mL Reagent I before use.

Reagent VI: Powder×4, store at -20°C . Add 208 μL distilled water before use. Mix thoroughly. Surplus solution store at 4°C .

Reagent VII: Liquid 250 μL×3, store at -20°C after spacing out. Avoid repeated freezing and thawing.

Reagent VIII: Liquid 12.5 μL×2, store at 4°C . Add 4 mL dissolved Reagent IV before use.

Reaction solution I: Add 7 mL Reagent I to Reagent II, heat slowly make it dissolve, add Reagent III after cooled. It can be prepared in two batches and determined.

Product Description:

Soluble Starch Synthase (SSS, EC 2.4. 1.21) usually present in the free matrix in the plastid matrix, which catalyzes the elongation of the starch chain, mainly responsible for the synthesis of amylopectin.

SSS catalyzes the reaction of ADPG with starch primer (glucan), transfers glucose molecules to starch primers, and simultaneously produces ADP. Add pyruvate kinase, hexokinase and 6-phosphate glucose dehydrogenase to the reaction system. These enzymes in turn catalyze NADP⁺ reduction to NADPH, the amount of NADPH produced is proportional to the amount of ADP produced in the previous step reaction, and the SSS activity can be calculated by measuring the increase of NADPH at 340 nm.

Required but not provided:

Ultraviolet spectrophotometer, water bath, desk centrifuge, transferpettor, 1 mL quartz cuvette, mortar/homogenizer, ice and distilled water.

Protocol:

I. Sample Preparation.

Add 1 mL Extract solution to 0.1 g tissue, homogenate on ice. 10000 g centrifuge at 4°C for 10 min. Take the supernatant on ice for test.

II. Determination procedure.

1. Preheat spectrophotometer for 30 min, adjust wavelength to 340 nm, set zero with distilled water.
2. Add reagents to centrifuge tube according to the following table.

Reagent Name (μL)	Test tube (V _T)
Sample	200
Reaction solution I	270
Mix thoroughly, keep warm for 20 min at 30°C, place at boiled water for 1 min (cover tightly to prevent water loss), cold on ice	
Reagent VIII	150
Mix thoroughly, keep warm for 30 min at 30°C , place at boiled water for 1 min (cover tightly to prevent water loss), cold on ice. 10000 g centrifuge for 10 min at room temperature. Take supernatant. Preheat Reagent V and supernatant at 37°C .	
Supernatant	450
Reagent V	300
Reagent VI	15
Reagent VII	15

Mix thoroughly. Measure the absorbance at 340 nm. Record the initial absorbance value A₁, after 2 min's reaction record absorbance value A₂. Calculate $\Delta A = A_2 - A_1$.

Note: If Reaction solution I had precipitation, mix thoroughly before add.

III. Calculation

1. Sample protein concentration

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the generation of 1 nmol of NADPH in the reaction system per minute every mg protein.

$$SSS (U/mg \text{ prot}) = [\Delta A \div (\epsilon \times d) \times V_T] \div (C_{pr} \times V_{SA} \div V_{RT} \times V_S) \div T = 43.2 \times \Delta A \div C_{pr}$$

This method needs to determine the protein concentration of crude enzyme solution.

2. Sample weight

Unit definition: One unit of enzyme activity is defined as the amount of enzyme catalyzes the generation of 1 nmol of NADPH in the reaction system per minute every g sample.

$$SSS (U/g \text{ weight}) = [\Delta A \div (\epsilon \times d) \times V_T] \div (W \div V_E \times V_{SA} \div V_{RT} \times V_S) \div T = 43.2 \times \Delta A \div W$$

V_T: Test volume, 0.78 mL.

V_{RT}: React solution volume, 0.62 mL .

V_E: Extract solution volume, 1 mL.

T: Reaction time, 20 min.

ε: The molar extinction coefficient of NADPH, 6.22×10^{-3} mL/(nmol · cm).

d: The optical path of cuvette, 1 cm.

V_{SA}: Sample volume, 0.2 mL.

V_S: Supernatant volume, 0.45 mL.

C_{pr}: Sample protein concentration, mg/mL.

W: Sample weight, g.

Experimental example:

1. Take 0.1g liver to 1ml extract solution, grinding on ice, 10000 rpm centrifuge at 4°C for 10 min, supernatant is ready for test, operate as the procedure, $\Delta A=A_2-A_1=0.394-0.211=0.183$, calculate content by sample weight: SSS (U/g weight)= $43.2 \times \Delta A \div W=79.056$ U/g weight.
2. Take 0.1g Ilex to 1ml extract solution, grinding on ice, 10000 rpm centrifuge at 4°C for 10 min, supernatant is ready for test, operate as the procedure, $\Delta A=A_2-A_1=1.31-1.303=0.007$, calculate content by sample weight: SSS (U/g weight)= $43.2 \times \Delta A \div W=3.024$ U/g weight.

References:

[1] Jiang H, Dian W, Wu P. Effect of high temperature on fine structure of amylopectin in rice endosperm by reducing the activity of the starch branching enzyme[J]. Phytochemistry, 2003, 63(1): 53-59.

Related products:

AK0364/AK0363 Bound Starch Amylosynthase (GBSS) Activity Assay Kit

AK0258/AK0257 Starch Branching Enzyme(SBE) Activity Assay Kit